



OCCURRENCE AND DISTRIBUTION OF ARBUSCULAR MYCORRHIZAL FUNGI FROM RHIZOSPHERE AND NON- RHIZOSPHERE SOIL OF SELECTED LOCALITIES OF PARNER TEHSIL

L. K. Dhumal¹ and R. K. Aher²

¹New Arts Commerce And Science College Ahmednagar, 414001, M.S. India

² New Arts Commerce And Science College Parner, 414302, M.S. India
lata_dhumal@rediffmail.com

Abstract:

Arbuscular Mycorrhizal (AM) species diversity and their root colonization patterns may vary in a plant species is influenced by environmental and biological factors. In the present study Rhizospheric and Non – Rhizospheric soils were collected from five localities of Parner Tehsil. Soil mycoflora was studied at vegetative stages of growth. Occurrence and distribution of Rhizosphere mycoflora was dominant over Non – Rhizosphere. The most Mycorrhizal fungi like *Acaulosporaelegans*, *A. foveata*, *A. laevis*, *Glomus aibidium*, *G. fasciculatum*, *G. globiferum*, *G. mosseae*, *G. versiformae* etc.

Keywords: Arbuscular Mycorrhizal, Rhizosphere and Non – Rhizosphere, Mycorrhizal root colonization, Parner.

Introduction

Frank (1885) first gave the name Mycorrhiza to describe the essential structure and functioning of the peculiar associations between the roots and ectomycorrhizal fungi. Arbuscular Mycorrhiza are considered as obligate biotrophic symbionts and are associated with the fine roots of over 80% terrestrial plant species. (Smith & Read 1997). In this plant fungus association, fungus depends upon host plant for nutrition and reproduction, and in return provides phosphates and essential mineral nutrients from soil to the host plant. Colonization by Arbuscular Mycorrhizal fungi assures good survival and growth of plants. (Vijaykumar and Abraham, 2001; Lakshaman & Patil, 2004) AM belongs to Zygomycete order Glomales.

The main function of AM fungi is of phosphorous transportation. Extra radical mycelium of AM fungi easily access P from soil and deliver to root cortical cells as polyphosphate which finally translocate to host plant after solubilization and it is estimated that external hyphae deliver up to 80% of P requirement of the plant (Matamoros et al 1999). Other than P translocation, AM fungi provides protection to the host plant roots from soil borne pathogenic attack they improve tolerance of plants to several abiotic stresses including drought, and saline stress condition by producing plant growth hormones (Evelin et al, 2009). In addition Mycorrhizal association also enhances nitrogen uptake as well as utilization of several micro nutrients AM successfully colonize with a wide range of plant species and are considered as non-host specific (Evelin et al 2009). Additionally different species

of AM fungi differ in their tolerance to adverse physical and chemical condition in soil (Kumar and Ghose, 2008).

In present study the mycoflora of Rhizospheric and Non – Rhizospheric soil of some selected localities of Parner Tehsil was studied.

Materials and Methods:

The Rhizospheric and non-Rhizospheric soil samples were collected from five selected areas of Parner Tehsil, areas like Takali Dhokeshwar, Wasunde, Khadakwadi, Padali Terfe Kanhur Pathar, and Hiware Korda. All the ten soil samples were brought to laboratory in sealed polythene bags. Then these samples were air dried in the shade at laboratory temperature. Then these samples were analyzed to its chemical parameters from the Agricultural Institute to examine its pH, salinity, organic carbon, Phosphorous, Pottassium, Copper, Iron, Zink & manganese.

One third part of each collected soil sample was used for AM spore extraction using the method described by Gerdemann & Nicolson (1963). In each case 50gm of sample was mixed with 500ml water followed by thorough stirring and then suspension was sieved through a series of sieves. Spores were collected on the filter paper and then counted using binocular compound microscope.

Results and Discussion:

AM isolated from Rhizosphere and Non-Rhizosphere soil of five different localities of Parner Tehsil are presented in Table No-1

In the Rhizospheric soil of 5 different localities 11-14 AM were isolated from vegetative stage. Non-Rhizospheric soil was also studied 7-8 AM were isolated from vegetative stage.

Total 20 species of Arbuscular Mycorrhizal fungi were isolated and identified from Rhizosphere and non Rhizosphere soil of five different localities from Parner Tehsil. It was clearly seen that species belonging to the three genera (Acaulospora, Glomus, Scutellispora) of AMF. Glomus species was most dominated than others two.

Table 1- Occurrence and distribution of Arbuscular Mycorrhizal fungi from Rhizosphere and non-Rhizosphere soil of selected localities of Parner Tehsil

AMF	Wasunde		Takali Dhokeshwar		Khadakwadi		Padalitarfe Kanhur Pathar		Hiware Korada	
	Rhizo soil	Non Rhizo	Rhizo soil	Non Rhizo	Rhizo soil	Non Rhizo	Rhizo soil	Non Rhizo	Rhizo soil	Non Rhizo
<i>Acaulospora foveata</i>	+	--	+	-	+	-	+	-	+	-
<i>Acaulospora laevis</i>	+	+	+	-	+	-	-	-	+	-
<i>Acaulospora elegans</i>	-	-	-	+	+	-	+	+	+	+
<i>Glomus aggregatum</i>	+	+	+	+	+	+	+	-	+	-
<i>Glomus albidum</i>	-	+	-	+	-	+	-	+	-	+
<i>Glomus constrictum</i>	+	-	+	-	-	-	-	+	+	-
<i>Glomus dimorphicum</i>	-	-	+	-	+	-	+	-	+	-
<i>Glomus etunicatum</i>	+	-	-	+	+	-	-	-	+	+
<i>Glomus fasciculatum</i>	+	+	+	+	+	+	+	+	+	+
<i>Glomus geosporum</i>	+	+	+	-	+	+	+	-	+	+
<i>Glomus globiferum</i>	+	-	-	-	+	-	+	-	+	-
<i>Glomus heterosporum</i>	+	-	+	+	-	-	+	+	-	-
<i>Glomus macrocarpum</i>	+	+	+	+	+	+	+	-	+	+
<i>Glomus monospermum</i>	-	-	-	-	+	+	-	-	-	-
<i>Glomus trimurales</i>	+	-	+	-	+	-	+	-	+	-
<i>Scutellispora dipurpurascens</i>	-	-	+	-	-	+	-	-	+	-
<i>Scutellispora aggregaria</i>	+	+	+	+	+	-	+	+	+	+
<i>Scutellispora heterogama</i>	+	-	+	-	+	-	+	-	+	-
<i>Scutellispora nigra</i>	+	+	+	+	+	+	+	+	+	+

Conclusions:

From the above observation table it was concluded that the species of Glomus were more diverse than the Acaulospora, Scutellispora. Glomus macrocarpum, G.fasciculatum, G.gigasporum was more dominated in both Rhizo as well as Non-Rhizospheric soil.

Acknowledgements:

The authors are thankful to Principle of New Arts Commerce and Science College, Ahmednagar, and Head of Department of Botany, New Arts Commerce and Science College Ahmednagar for providing necessary facilities for this work. The authors are also thankful to Principle of New Arts Commerce and Science College, Parner for encouragement and support.

References:

- Abbott LK, Robson AD, 1984. The effect of VA mycorrhizae on plant growth. In: Powell CL, Abbott LK, Robson AD, 1991. Factors influencing the occurrence of vesicular-Arbuscular mycorrhizae. Agr. Ecosys. Environ. 35: 121-150.

Aliasgharzadeh N, Saleh Rastin N, Towfighi H, Alizadeh A, 2001. Occurrence of Arbuscular Mycorrhizal fungi in saline soils of the Tabriz plain of Iran in relation to some physical and chemical properties of soil. Mycorrhiza 11: 119-122.

Bagyaraj DJ. (ed.) Mycorrhizae. pp. 113-130. Boca Raton, Florida, USA: CRC Press.

Bagyaraj D.J., 1984. Biological interactions with VA Mycorrhizal fungi. In: VA Mycorrhiza (Ed. Powell, C.L. and Bagyaraj, D.J.) CRC Press, Florida. pp. 131-154

Brundrett M. (2004). Diversity and classification of Mycorrhizal associations. *Biol Rev*, 79: 473-495

David, M. Sylvia. And Jacob, N. Burks. (1988). selection of a vesicular-Arbuscular Mycorrhizal fungus for practical inoculation of Uniolapaniculata. *Mycologia*, 80(4): 565-568.

Gerdemann JW and Nicolson TH., 1963. *Int. J. Curr. Microbiol. App. Sci* (2014) 3(6): 527-539

538 Spores of Mycorrhizal Endogone species extracted from soil by wet sieving and decanting. *Trans Br Mycol Soc.* 46(2):235-244.

Gianinazzi-Person, M. and Gianinazzi, V. (1994). Biodiversity in Arbuscular fungi. *Mycol Res*, 98:705-715.

- Giovannetti M, Mosse B (1980) An evaluation of techniques for measuring vesicular ArbuscularMycorrhizal infection in roots. *New Phytol* 84:489–500
- Harrison RW (1955) A method of isolating vesicular-Arbuscularendophytes from roots. *Nature* 175:432-159
- Johnson, D., Booth, R. E., Whiteley, A. S., Bailey, M. J., Read, D. J., Grime, J. P. And Leake, J. R. (2003). Plant community composition affects the biomass, activity and diversity of microorganisms in limestone grassland soil. *Eur. J. Soil Biol*, 54: 671–677.
- Marschner, H. and Dell, B. (1994). Nutrient uptake in Mycorrhizal symbiosis. *Plant soil*, 159: 89-102.
- Morton JB, Benny GL (2001) Two new families of Glomales, Archaeosporaceae and Paraglomaceae, with two new genera Archaeospora and Paraglomus, based on concordant molecular and morphological characters. *Mycologia* 93:181–195
- Morton, J. B. (2001). Taxonomy of VA Mycorrhizal fungi: Classification, nomenclature, and identification. *Mycotaxon*, 32: 267-324.
- Philips, J. H. and Hayman, D. S. (1970). Improved procedures for clearing roots and staining parasitic and vesicular – ArbuscularMycorrhizal fungi for rapid assessment of infection. *Transactions of the British Mycological Society*, 55: 158-161.
- Scheussler, A., Schwarzott, D. and Walker, C. (2001). A new fungal phylum, the *Glomeromycota*: phylogeny and evolution. *Mycol. Res*, 105: 1413-1421
- Smith, G.W and Read, D.J. (1997). Mycorrhizal symbiosis. Academic Press, London, UK. Pp.432.
- Srivastava, D., Kapoor, R., Srivastava, S.K. and Mukerji, K.G. (1996). Vesicular ArbuscularMycorrhizal-an overview. In: Mukerji, K.G. (eds.) *Concepts in Mycorrhizal Research*. Kluwer Academic Publishers, Netherlands, pp: 1-5.