ANNUAL MORPHOLOGICAL VARIATION IN FEMALE REPRODUCTIVE SYSTEM OF SPOTTED SNAKEHEAD _CHANNA PUNCTATA_ (BLOCH, 1793)

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Abstract:
Snakehead _Channa punctata_ (Bloch, 1793) is popular food fish. Reproductive cycle or breeding cycle of _C. punctata_ has five phases viz., resting phase, early maturing phase, prespawning phase, spawning phase and spent phase. Gonadosomatic index (GSI) reveals the period and duration of different phases in reproductive cycle. GSI is maximum during spawning phase while it is minimum in resting phase. Snakehead, _C. punctata_ is annual breeder.

Ovary of _C. punctata_ increases its size and shape from resting phase to spawning phase and again reduces its size in spent phase. In different phases of reproductive cycle ovary also changes its color. Resting phase ovary is whitish which become pinkish during early maturing and prespawning phase due to vascularization of ovary. It become transparent and yellowish due to thinning of tunica albuginea and underlying yellowish eggs. Smooth contour of ovary become granular during spawning phase.

Key words: _Channa punctata_, Gonadosomatic index.

Introduction:
Spotted snakehead, _Channa punctata_ (Bloch, 1793) is highly esteemed as food (Chakrabarti, 2006). All the species of the _Channa_ viz., _C. punctata_, _C. marulius_, _C. striata_ and _C. gachua_ bear a pair of folded sac like outgrowth of pharynx called pharyngeal outgrowth to store air (Chakrabarti, 2006). This pharyngeal outgrowth provides these fishes a tenacity to survive outside the water for considerable period. Reproductive histology can be well investigated by various histological techniques. Ovarian cycle and spawning season of _Ophiocephalus punctatus_ inhabiting Jammu water was documented (Malhotra et al., 1978) where six stages of oocytes were reported by following method of Srivastava and Rathi (1970). Variation in gonadal cycle, spawning time and spatial behaviour of this fish were well documented (Swaroop, 1954; Belsare, 1962; Malhotra et al., 1978).

Present study was undertaken to ascertain the breeding cycle, spawning season and to study ovarian morphological changes in _C. punctata_.

Materials and Methods:
Matured female _C. punctata_ of 150-200 gm weight were collected/procured from various water bodies of eastern Vidharbha. Fishes were acclimatized in aquaria for one week. After sacrificing fishes, ovaries were dissected out and fixed in Bouin's fluid. After fixation, ovaries were cutted in smaller pieces and cutted pieces of tissues were washed and transferred to 70% alcohol and dehydrated in graded alcohol, cleared in xylene and embedded in paraffin wax at 60°C- 62°C. Blocks of tissues were trimmed and serial sections of these blocks were cut on Cambridge (rocking) microtome at 6-8 µm thickness in transverse plane. The sections were fixed on clean slide and later stained by Haematoxyline- Eosine procedure (Lillie, 1965). Sections were observed under Karl Zeiss microscope and photographed by photographed by Tucsen USB 2.0 H series.

Gonado-somatic index (GSI) was used to ensure the different phases of reproductive cycle. GSI was calculated by formula:

\[
\text{GSI} = \left( \frac{\text{Wg} \times \text{W}^{-1}}{\text{W}} \right) \times 100
\]

Where:
Wg- Weight of gonads (g), W- Body weight (g).

Results were expressed as the mean and standard error of mean (SEM). Difference between means were analysed by one-way ANOVA and student T-test. Level of significance was set at \( P \leq 0.05 \).

Morphological study of ovary was studied after giving midventral cut to abdomen of fish. Dissected fish were photographed by Nikon camera.

Results:
Paired ovary of _C. punctata_ is suspended from dorsal wall of peritoneal cavity by mesorchium. Two ovarian lobes in _C. punctata_ are of unequal length. The ovarian lobes are of smooth cyclindrical in shape. The ovarian lobes on outer side covered by tunica albuginea (Fig. 1a).

Based on GSI, morphology of the ovarian lobe and proportion of oocytes present in ovarian lobes, five phases of maturation or reproductive cycle of _C. punctata_ were identified.

1. Resting phase (Mid December- February)
2. Early maturing (March- April)
3. Prespawning (May)
4. Spawning (June- October)
5. Spent (November- Mid December)

GSI in resting phase is \( 0.58 \pm 0.04 \) (Table 1, graph 1). In resting phase, ovary is slender and whitish (Fig.1a). Histologically in resting phase,
The ovary shows distinguished ovigerous lamellae hanging in ovocoel (Fig. 2a). Ovigerous lamellae possess primary oogonia, chromatin nucleolus oocyte, early perinucleolar oocyte and few late perinucleolar oocytes (Fig. 2a).

Early maturing phase ovary slightly swollen and become pinkish (Fig. 1b). Besides primary oogonia, chromatin nucleolus oocyte, early perinucleolar oocyte and late perinucleolar oocyte; vitellogenic oocyte also seen. GSI is early maturing phase increases slightly and is 1.99 ± 0.9 (Table 1, graph 1).

Prespawning ovarian lobe further enlarges become transparent and vascularised (Fig. 1c). Histologically it shows few additional ripe eggs besides the oocytes stage which were seen in early maturing phase (Fig. 2c). GSI of prespawning ovary increases further and is 4.49 ± 0.89 (Table 1, graph 1).

In spawning phase, ovary attains maximum size and outer tunica albuginea become thinner and completely transparent. Smooth contour of ovary look granular due to presence of bulging oocytes (Fig. 1d). In this phase, ovary become highly vascularized (Fig. 1d). Histologically ovary show abundant ripe eggs along with other stages of oocytes (Fig. 2d). Maximum GSI attained during spawning phase and is elevated to 14.49 ± 0.73 (Table 1, graph 1).

In spent phase, ovary become flaccid and thinner (Fig. 1e). During this phase, follicles expel the oocyte thus ovary showed several ovulated follicles. Several atretic follicles were also reported during this phase (Fig. 2e). GSI again lowered down and is 0.60 ± 0.02 (Table 1, graph 1).

**Table 1: GSI in various phases of breeding cycle.**

<table>
<thead>
<tr>
<th>Phases</th>
<th>GSI</th>
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<tbody>
<tr>
<td>Resting phase</td>
<td>0.58 ± 0.04</td>
</tr>
<tr>
<td>Early maturing phase</td>
<td>1.99 ± 0.9</td>
</tr>
<tr>
<td>Prespawning phase</td>
<td>4.49 ± 0.89</td>
</tr>
<tr>
<td>Spawning phase</td>
<td>14.49 ± 0.73</td>
</tr>
<tr>
<td>Spent phase</td>
<td>0.60 ± 0.02</td>
</tr>
</tbody>
</table>
Discussion:

In *C. punctata*, a pair of elongated ovaries was located in coelomic cavity. Each ovary leads into short oviduct which later opens outside through urinogenital opening. Ovaries from resting phase to spawning phase turn whitish to yellowish in early maturing phase and then become transparent in spawning phase. In spent phase, it looks whitish. Similar result were reported in *Carassius auratus* (Munkittrick and Leatherland, 1984), *Anabas testudineus* (Bahera, 2015). Ovary attain maximum size during spawning phase. Similar observations were describe in several cyprinid fishes (Al-Daham and Bhatti, 1979; Al-Nouri, 1996; Bardakci et al., 2000).

Symmetry wise, ovaries could be categorized into symmetric and asymmetric ovaries. Most of the teleostean species exhibit symmetric ovary with similar ovarian lobe. Some teleost show remarkable difference in the length of ovarian lobes. Such ovary represents asymmetric ovaries. *Anchoa mitchilli* shows asymmetric ovaries (Kobelkowsky, 2012). To ascertain type of *C. punctata* ovary is difficult task as difference between right and left lobe of ovary is less during the resting and early maturing phase, difference between two lobe became remarkable in spawning phase.

GSI is strongly correlated with gonadal development and maturity of fish (Rae and Calvo, 1995; Zimmerman 1997; Koya et al., 1998). Maximum GSI was reported during spawning phase. Similar reports were describe in *Cynoglossus arel* and *C. lida* (Rajguru, 1992), *Potagonotothen tesselata* (Rae and Calvo, 1995) and in rainbow trout, *Oncorhynchus mykiss* (Sharma and Bhat, 2014).

Ovary of *C. punctata* is asynchronous type as each phase of breeding cycle shows different type of oocytes. Similar asynchronous ovary was reported in *Chirostoma hermodictianum* (Cardenas et al., 2008).

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