



THE DEVELOPMENT OF MEGASPORANGIUM, MEGASPOROGENESIS AND FEMALE GAMETOPHYTE, EMBRYOGENY AND CARYOPSIS IN *ARUNDINELLA PUMILA* (HOCHST.EX.A. RICH)

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ABSTRACT:

The paper deals with embryological investigation of *Arundinella pumila*. The ovules are bitegmic, pseudocrassinucellate and campylotropous, polygonum type embryo sac with antipodal complex. The embryo development conforms to Asterad type. Endosperm, caryopsis, seed coat and pericarp, have been described critically.

Key words: - Poaceae, Poodeae, Hypostase, Embryology, Embryo development, Caryopsis.

INTRODUCTION:

The division of the Poaceae into two subfamilies viz., Pooideae & Panicoideae as proposed by Brown (1814) is being maintained even today. Embryological features of this taxon follows the uniform pattern at subfamily level (Narayanswami 1955 a,b,c, 1956; Koul, 1997 a,b; Raju 1980; Bhanwara et al., 1991; Deshpande & Makde 1994; Nikhade & Makde, 1997). Caryopsis a unique fruit of Poaceae were pericarp free from seedcoat. The present paper pointed out the some embryological character of taxonomic value.

MATERIALS AND METHODS:

The material of *Arundinella pumila* was collected from Totaladoh localities of Nagpur (M.S.) India. The voucher specimens (N-7) submitted to the herbarium Department of Botany RTM Nagpur university Nagpur. Spikelet's at various stages of development were fixed in 70% F.A.A. Customary methods of dehydration, infiltration and embedding followed. Sections were cut 8-12 μ thick & stained with Delafields hematoxylin.

Erythrosin and light green stain was used as counterstain. Mature embryos were dissected out and stained with safranin.

Ovule, Megasporogenesis and female gametophyte.-

The family is characterized by bi/tricarpellary, syncarpous pistil having a single ovule in the unilocular ovary (Lawrence, 1951, Hutchinso, 1973). The ovules are campylotropous, tenuinucellate & bitegmic (Fig. 1M-N). The integuments both inner & outer are two layered for their major portion. The hypodermal female archesporium functions directly as megaspore mother cell (Fig. 1S). The megaspore mother cell undergoes meiosis I followed by successive cytokinesis. Meiosis II in the dyad cells is synchronous. Thus, megaspore tetrads are formed at this stage both the integument are well developed and these extend almost upto the shoulder at the nucellus. The chalazal megaspore alone functions & remaining three megaspore degenerate (Fig. 1Q). The degenerating remains persist up to the organsiation of mature embryo

sac (Fig. 2C). The mature embryo sac more or less cylindrical. The egg apparatus consists of a flask shaped egg & two synergids. The two polars are quite close approximation to each other very near the egg (Fig. 2C). They fuse to form secondary nucleus just prior to fertilization (Fig. 2D). At the time of embryo sac organization there are only three antipodals cells located at chalazal end, later on mitotic division in them results in an increase in their number. Thus, antipodal complex composed of 10-12 cells (Fig. 2D).

Hypostase :

A group of nucellar cells at chalazal end of the ovule becomes conspicuous & differ from the adjoining cells termed as hypostase. The hypostase is well developed & cylindrical. Its cells get filled with tanniniferous granular deposits at the globular stage of embryo development (Fig. 2E). At advanced stage of embryo development they are completely packed with tannin (Fig. 2F).

Endosperm:

The endosperm development is free nuclear type initially. It later becomes cellular & shows deposition of reserve food matter. At the bicelled pro-embryonic stage there are 12 nuclei & they are arranged in a peripheral cytoplasmic layer around a central vacuole (Fig. 3G). When the embryo enter 3rd & 4th cell generation, there are sudden increase in the numbers of free endosperm nuclei (Fig. 2H). Initiation of the cellular phase in endosperm commences first in the micropylar region around the embryo & gradually proceeds towards the chalazal end. The cell wall formation begins at the periphery and progress centripetally (Fig.2J)

At the advanced globular embryo stage, endosperm becomes completely cellular with a single aleurone layer (Fig. 2I). During later stage of embryo development starch grains accumulation takes place in this tissue. The embryo consume only a part of cellular endosperm in the micropylar region, rest of the

part occupied by cellular endosperm (Fig. 2K). Thus, the seeds are endospermic.

Embryo Development:

The transverse division of the zygote results in the cells ca & cb (Fig. 3A). The cell cb divides transversely forming m & ci (Fig. 3B,C). The pro-embryonic tetrad produced at the end of 2nd cell generation can safely be described as T-shaped or assigned to series A2 of Soueges (1951) system of classification. During the 3rd cell generation sequential stage could not get. In the 4th cell generation tier q definitely gets demarcated into tiers l & l' Fig. 3D,E). The tier ci divides transversely & vertically into n & n' tier. The tier o & p are the derivatives of n' & forms a suspensor.

During further development tiers l & l' and m do not follow a regular sequence but occur in a different plane results in a massive globular embryo (Fig. 3F,G). The flaps of the coleoptile & stem tip, arise as bulges on one side of the embryo (Fig.3H, I). The entire tier l gives rise to the single cotyledon and cleft (upper lip of the coleoptile) organized. The tier l' contributes for the stem tip, first leaf & lower lip of the coleoptile respectively (Fig. 3L). In mature embryo pro-vascular strand joined to the plumule radicle axis (Fig. 3M). The plumule (shoot apex) is laterally disposed & enclosed by coleoptile. The shoot apex is dome shaped covered by few primordial leaves (Fig, 3N). At the radicular end there is a distinct root cap with coleorhiza covering. The mature embryo is not embedded in the endosperm but remains peripheral to it.

Seed coat and Pericarp:

The ovules are bitegmic, both the integuments are two layered. At the mature embryo sac stage both the layer of outer integument get highly stretched & gradually start degenerating (Fig. 4B). The inner integument though persists at the earlier stage, its outer layer starts collapsing at globular embryo stage, (Fig. 4C). At advanced embryo stage outer layer of inner integument

disorganizes & cells of the inner layer get filled with tannin (Fig, 4D). Thus at maturity only inner layer of inner integument persist as a seed coat. (Fig. 4E). Pericarp 4-6 layers including the outer & inner epidermis layers remain constant at mature stage of the grain. (Fig. 4E). Pericarp remains free from the seed coat. The cells of the outer epidermis show thickened inner tangential walls with tannin deposition (Fig. 4E).

RESULT & DISCUSSION:

The present embryological work on *Arundinella pumila* which belongs to subfamily, Pooideae resembles in most of the embryological features of the family work out by earlier worker.

The ovules are campylotropous, bitegmic tenuinucellate & shows the feeble tendency towards Pseudocrassinucellar. The polygonum type of embryo sac development (Maheshwari, 1950). This is a very constant feature in the family investigated so far (Anderson, 1927; Beck & Harten, 1932; Chandra, 1963b, 1970; Venkateshwarlu & Devi 1964; Raju, 1980; Febulaus & Pullaiah, 1992). The three celled antipodal & antipodal complex at chalazal end in the female gametophyte in this taxon. In the Pooideae they are lateral (Chandra, 1963b; Venkateshwarly & Devi 1964; Davis, 1964; Bhanwra, 1988). Therefore, the position of antipodals has not been taken as a taxonomic character. The chief function of the hypostase is the translocation of nutrients; it may acts as a secondary storage tissue. This observation is supported by the findings of present paper. The endosperm development is free nuclear earlier but later on it becomes cellular at the end of globular or advanced embryo phase & the meristematic activity in peripheral layer of endosperm which forms a characterisitic feature of the family (Narayanswami, 1953, 1955a,b,c, 1956; Chandra, 1963; Sapre, 1964; Johri & Ambegaonkar, 1976.)

The embryo development conforms to the Asterad type of Johansen (1950). According to Soueges

(1951) the development conforms to grand period I, series A, subseries A2 & megarche type II. The creation of a new variation viz, 'Graminad Type' on the basis of oblique plane during pro-embryonic development by Batygina (1969a,b). This is criticised by Gerassimova-Navaschina (1977).

The fruit in grasses is classified under Caryopsis and has been define as seed. The testa and pericarp are further reported to be fused with each other (Anderson, 1927, Julien and Aldama , 1939). However data available from later studies on grasses do not agree with these findings (Narayanswami, 1955 a, b, c; Raju, 1980; Bhanwara, 1988; Ghaisas, 1991; Despande and Makde, 1994; Nikhade and Makde, 1997). The present study also testifies to this. Sendulsky et. Al. (1986) stated that caryopsis essentially is dry indehiscent fruit where pericarp is free or closely adherent to seed coat the author feels that in the light of theses finding there is an urgent need to redefine the term caryopsis.

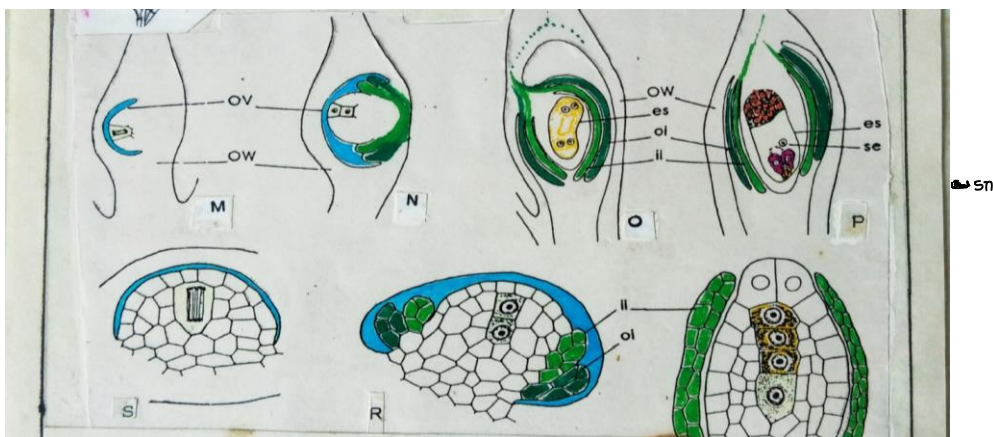
REFERENCES:

- Anderson, A, M, 1927, Development of the female gametophyte and caryopsis in *Poa pratensis* & *poa compressa*. J. Agric., Res. 34:1001:1018.
- Batygina, T.B. 1969 a. Embryogenesis in the genus *Triticum Linn.* as related to the problem of monocotyledony and remote hybridization in Gramineae. Bot. Zh., 53.:480-490.
- Batygina, T.B. 1969 b, "On the possibility of a new type of embryogenesis in angiosperm." Rev. Cytol Boil.Veg. 32:335-341.
- Batygina T.B. 1974. Wheat embryology "kolas" Leninggard (In Russian).
- Bhanwara R.K.1985.Embryological Studies in five species of *Eragrostis* Beauv.(Gramineae) Res.Bull(Sci) of Punjab Univ,ed,(Parts-I-II):17-23.

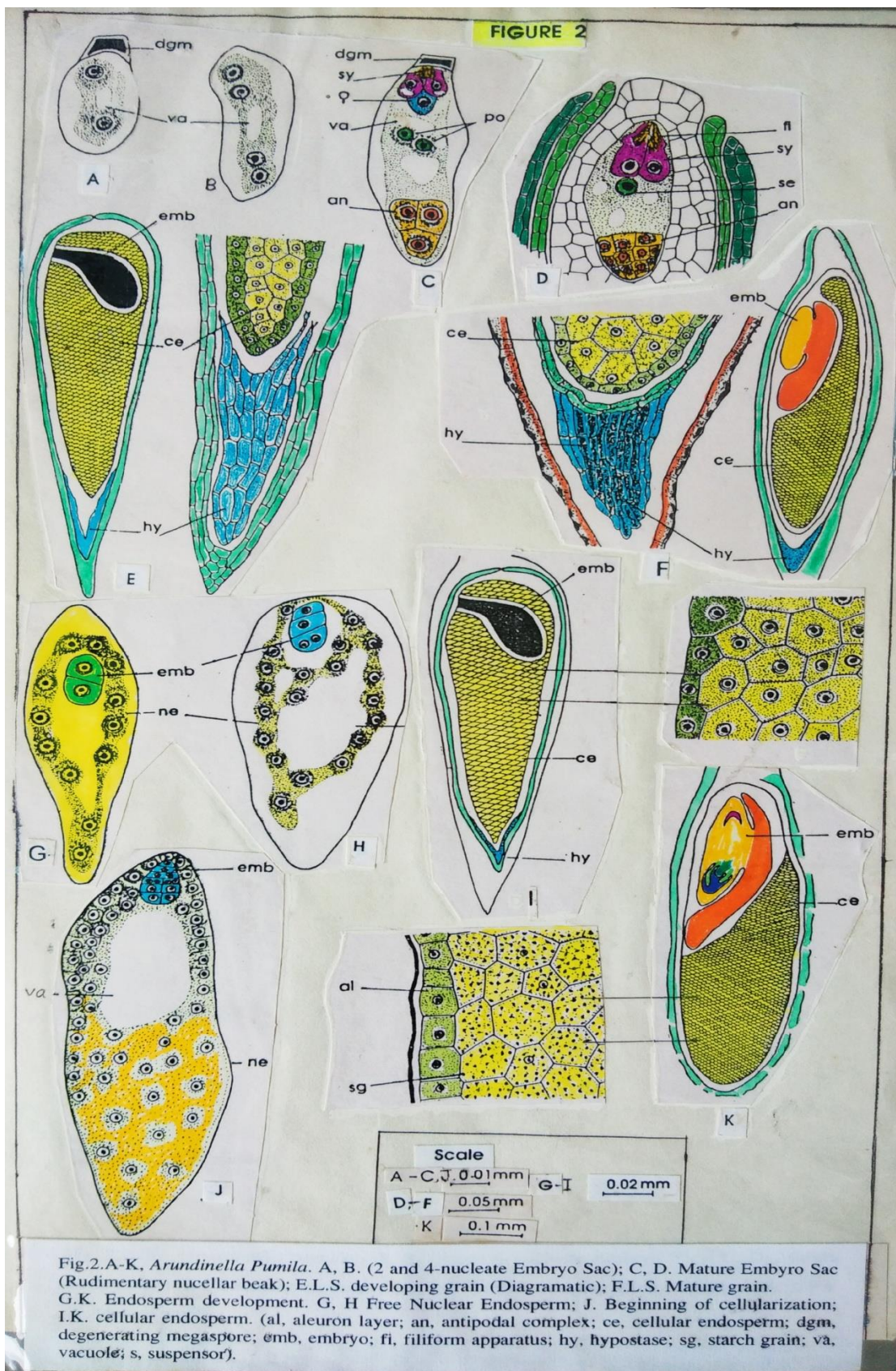
- Bhanwara R.K.1988 Embryology in relation to systematics of gramineae. Annals of botany. 62:225-233.
- Bhanwara. R.K., N. Kaur & A Garg. 1991 Embryological studies in some grasses and their taxonomic significance. Bot. J. Linn. Soc 107:405-419.
- Beck, P & J.S, Horton. 1932. Microsporogenesis & embryology in certain species of *Bromus*, Bot. Gaz, 93:42-48
- Bhaskute, Shshma M. In vitro studies on *Dendrocalamus strictus* Nees & *Bambus arundinacea* Moon with some observations on their embryology & histochemistry. Ph.d thesis Nagpur university Nagpur.
- Brown, R. 1814. General remarks, geographical and systematical on the botany of *Terra Australis* Pb 533-613 in M Flinder. A voyage to *Terra Australis*. Vol 2 london.
- Chandra, N. 1963 a. Morphological studies in gramineae IV Embryology of *Elusine indica* & *Dactylocaenium aegypticum*. Proc. Indian Acad Soci B 58:117-127.
- Chandra, N 1963 b. Some Ovule characters in systematics of the Gramineae. Curr Sci., 32:277-279.
- Chikkannaiah, P. S. and M. S. Mahalingappa 1976 a. Embryological studies in *Elusine*. Abst. In embryology of crop plants. Indo-soviet symposium sponsored by INSA and University of Delhi P. 7.
- Chikkannaiah, P. S.S and M.S.Mahalingappa 1976 b. The female gametophyte and activities of antipodals in *Cynodon dactylon* (L.). Pers Abst. In Physiocoty of sexual reproduction, International symposium Ludhiana .P 62.
- Christenson, J. E., H. T. Horner and N.R. Lersten. 1972. Pollen wall and tapetal orbicular wall development in *Sorghum bicolor*. Am .J. Bot. 59:43-52.,
- Devis, G. L. 1966. Systematic Embryology of the Angiosperms. Wiley, New York.
- Diwanji , V. B.1976. "Embryological studies in Gramineae" Ph.D. Thesis, University of Indore., Indore.
- Deshpande, P.K. 1965. Development of embryo & endosperm in *Eragrostis uniolooides* (Poaceae) Plant syst Evol., 125:235-259.
- Deshpande, P.K. & K.H. Makde, 1994. Embryo & fruit in the Poaceae. Advances in Plant Reproductive Biology Vol 1 Eds Chauhan & Panday Narendra Publishing House, Delhi : pp-101-115.
- Febulaus, G.N.V. & T. Pullaiah. 1990, Embryology Of *Chloris roxburghiana* Shult (Poaceae).J.Indian Bot. Soc Vol. 69:52-53.
- Gerassimova-Navashina, E.N. 1972, Development determination of the embryo structure In angiosperms. Bot. Zhur. 57(4): 441-457.
- Ghaisas, V.A. 1991: Morphological & Histochemical Investigations on some oil yielding Grasses. Ph.D. Thesis Nagpur Univ. Nagpur.
- Hutchinson, J. 1073. *The families of flowering plants*, 3rd edn., Clarendon, Oxford.
- Johansen,D.A. 1950: Plant Embryology. Waltham, Mass. U.S.A.
- Johri, B.M & K.B. Ambegaonkar, 1976. Seed Development in Triticales Phytomorphology 25:112-117.
- Koul, A.K. 1970 a, Cytoembryological Studies in oriental Maydeae I; *Coax acuatica* Roxb, Proc., Nat, Acad , Sci, 40:163-178,
- Koul, A.K. 1970 b, Cytoembryological studies in oriental Maydeae II; *Chionachne koenigii* Thu. Ibid, 40:178-190.
- Lawerence, G.H.M. 1951. *Taxonomy of vascular plants*. Mac Millan, New York.
- Maheshwari, P. 1950, An Introduction to the embryology of Angiosperms, Mc Grew Hill., New York.

- Makde, K.H. 1973, Embryological and Palynological studies in the Cyperaceae. Ph.D. Thesis, Nagpur University Nagpur.
- Narayanswami, S. 1953, The structure and development of Caryopsis in some Indian Millets. I. *Pennisetum typhoideum* Rich. Phytomorph. 3:98-112.
- Narayanswami S. 1955a, The structure & development of the Caryopsis in some Indian Millets III. *Panicum millare* Lamk. & *P. miliaceum* Linn. Liodyia. 18:61-73.
- Narayanswami, S. 1955 b, The structure & development of the Caryopsis in some Indian Millets. IV. *Echinochloa frumentacea* Linn. Phytomorphology. 5:161-171.
- Narayanswami. S. 1955c. The structure & development of the Caryopsis in some Indian Millets. V. *Eleusine coracana* Gaertn. Mich. Acad. Sci. A.L., 40:33-46.
- Narayanswami, S. 1956, Structure & development of the Caryopsis in some Indian Millets. VI *Setaria italic*. Bot. Gax. 118:112-122.
- Nikhade, C.A. & K.H. Makade, 1997, A contribution to the embryology of *Perotis indica* (L.) O.Ktze. J. Natl. Bot. Soc; 51:33-41. India.
- Padhye. M.D. & A.G. Untawale, 1967. Embryological and taxonomical studies in the Cyperaceae with some observation on the embryology *Passiflora foetida*. Ph.D. Thesis Nagpur Univ. Nagpur.
- Raju. P.S G. 1980. Embryological & histochemical studies of some Crop plants (Gramineae) Ph.D. Thesis, Nagpur University Nagpur.
- Sapre, A.B. 1976. Embryology of a multicaryotic variety of Rice Abst. In Embryology of crop plants. Indo-Soviet symposium, Sponsored By INSA & Univ, Of Delhi P.36.
- Soueges, R. 1951. Embryogenic et classification 4. Essaid unsystem embryogonique, Paris, Hermiann & Cie.
- Untawale, A.G. 1970. Embryological studies in Cyperaceae. Ph.D. Thesis, Nagpur Univ. Nagpur.
- Venkateshwarlu. J&P. I. Devi. 1964. Embryology of some Indian grasses. Curr. Sci. 33:104-106.

FIGURE 1



Fig, 1M-S; Megasporogenesis & Female Gametophyte Development. M-P. L.S. ovary (diagrammatic) Development of campylotropous ovule. (es, embryo sac ; sn, secondary nucleus ; ii, inner integument ; oi, outer integument ; ov, ovary; ow, ovary wall;)



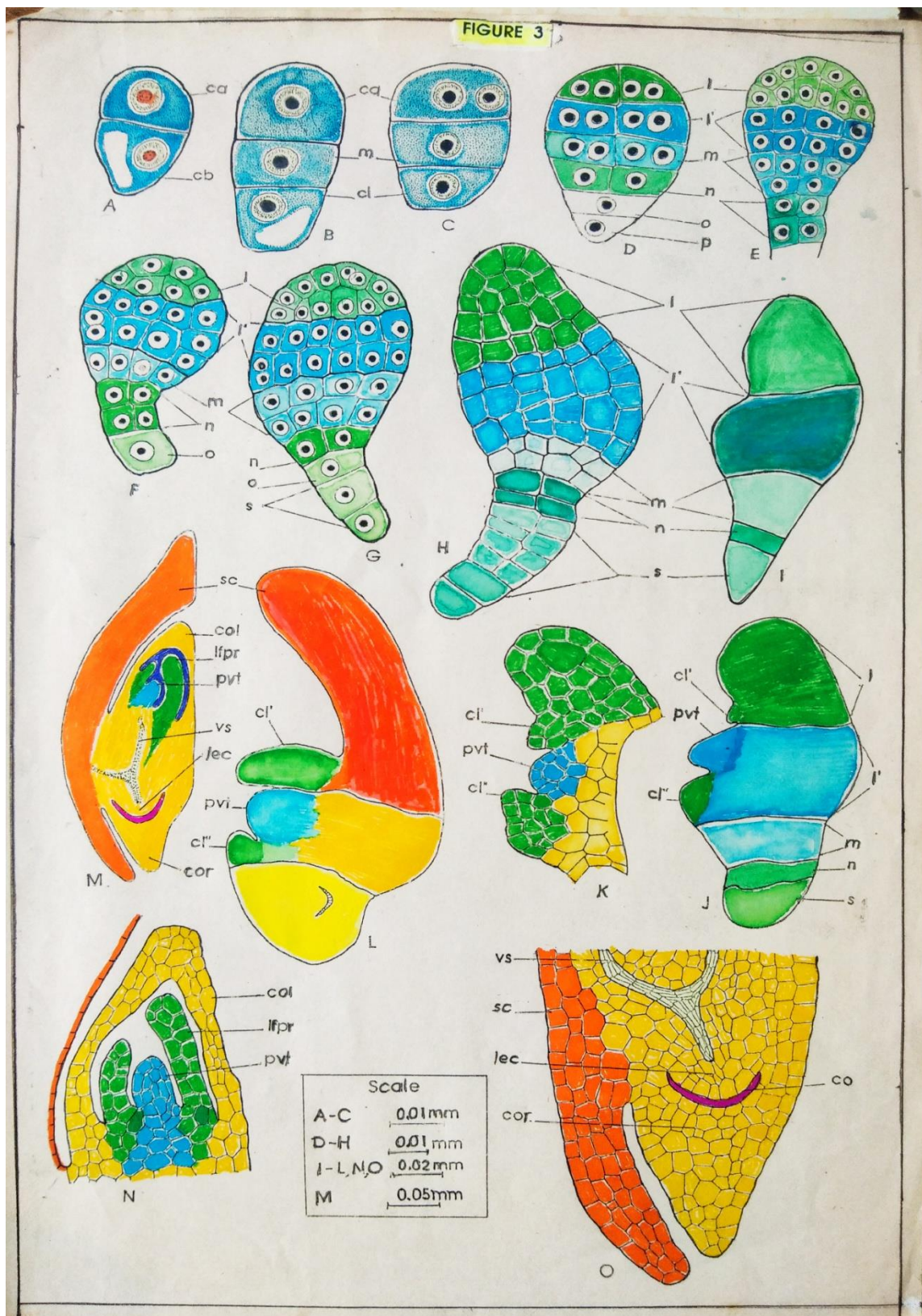


Fig.3 A-O-Stages in the development of embryo. (co, root cap; col, coleoptile; cor, coleorhiza; cl' cleft one; cleft two; iec, initial of the root cortex; lfpr, leaf primordia; pvt, stem tip; sc, scutellum; s, suspensor).

