



# Designing a Urease Biosensor Using Modified Electrode with Pb doped Al<sub>2</sub>O<sub>3</sub> Nanoparticles

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## Abstract

Chitosan (CH) -Pb doped Al<sub>2</sub>O<sub>3</sub>nanoparticles-nanocomposite film has been fabricated onto gold plate to immobilize urease (Ur) via physisorption for urea detection. Both CH-Pb doped Al<sub>2</sub>O<sub>3</sub>/gold electrode and Ur/ CH- Pb doped Al<sub>2</sub>O<sub>3</sub> /gold bioelectrode have been characterized using cyclic voltametry (CV) and electrochemical impedance spectroscopy (EIS). Cyclic voltametry shows the persence of Pb doped Al<sub>2</sub>O<sub>3</sub> nanoparticles results in increased electroactive surface area of CH-Pb doped Al<sub>2</sub>O<sub>3</sub>nanobiocomposite for immobilization of enzyme (Ur), enhanced electron transfer between Ur and electrode and electrochemical impedance spectroscopy (EIS) shows the conductivity of composite electrode.

**Keywords:** Biosensor; Pb doped Al<sub>2</sub>O<sub>3</sub>; Urease (Ur); Nanoparticles; Chitosan (CH)

## Introduction:

In the past few years, substantial research effort has been devoted to the development of organic biosensors, since they combine general merits of electronic sensors, like speed, size and system integration, with the ones of organic semiconductors, such as low cost production, facile integration with flexible substrates, and biocompatibility [1,2]. According to the fact, the design of modern biosensors is strictly combined with achievements of the nanotechnology. Nanostructures and nanotechnology are connected with design and production of material and devices in range 1-100 nm. In this context, using of different type of nanoparticles or nanosized materials could improve i.e. sensitivity of device on the step of this construction already [3-5].

The increasing demand for clinical diagnostics relating to kidney and liver diseases has necessitated evolution of new methods for faster and accurate estimation of urea in desired samples including urine and blood samples. The increased urea concentration (normal level in serum is 8-20 mg/dL) causes renal failure (acute or chronic), urinary tract obstruction, dehydration, shock, burns and gastrointestinal bleeding. Moreover, decreased urea concentration causes hepatic failure, nephritic syndrome, cachexia (low-protein and high-carbohydrate diets) [6-8]. The conventional methods for urea detection including gas chromatography, calorimetry and fluorimetric analysis suffer from complicated sample pretreatment and are unsuitable for on-line monitoring. Electrochemical biosensors have been considered to provide interesting alternatives due to their simplicity, low cost and high sensitivity [8, 9].

More recently, biosensors have emerged as a promising technology, especially for applications requiring rapid and continuous monitoring. Biosensors are being applied to a wide variety of analytical problems such as in medicine [10-





12], environment [13–15], food and process industries [16,17], security and defence [18]. Although urea biosensors emphasising on better sensitivity or higher response range [19] are reported, not much effort has been made in resolving the drawbacks of enzyme instability, difficulty in storage and handling, and fragility of the immobilization matrix.

Immobilization of Ur onto a suitable matrix is a crucial for the development of an electrochemical urea sensor [1-4]. In hybrid nanobiocomposites, surface functionalization of nanoparticles allows their covalent attachment and self-assembly on surfaces that can be used for loading of desired biomolecules in a favorable microenvironment for development of a biosensor [20, 21]. In this context, metal oxide nanoparticles-chitosan (CH) based hybrid composites have attracted much interest for development of a desired biosensor [20].

In this manuscript, we report results of studies relating to the immobilization of Ur onto CH-Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite film.

### **Experimental details**

#### **Material:**

All chemicals and solvents were analytical grade and purchased from commercial sources.

#### **Preparation of Pb doped $\text{Al}_2\text{O}_3$ nanoparticles**

The Pb doped  $\text{Al}_2\text{O}_3$  nanoparticles prepared by using sol-gel citrate method. The stoichiometry mixture of lead nitrate, aluminium nitrate magnetically stirred with citric acid and ethanol at  $80^\circ\text{C}$  for 3hrs to get homogeneous and transparent solution. The solution was further heated at about  $130^\circ$  for 12 hrs. in pressure vessel. The prepared product was subjected to 3hrs heat treatment at  $350^\circ\text{C}$  in muffle furnace and then milled to a fine powder. The dried powder then calcinated in range of  $350^\circ$ - $650^\circ\text{C}$  in order to improve the crystallinity, sensitivity and selectivity of material.

#### **Preparation of CH-Pb doped $\text{Al}_2\text{O}_3$ /Gold nanobiocomposite electrode**

Pb doped  $\text{Al}_2\text{O}_3$  nanoparticles prepared using sol-gel method are dispersed into 10 mL of CH (0.5 mg/mL) solution in acetate buffer of 0.05M at pH 4.2 under continuous stirring at room temperature. Finally, viscous solution of CH with uniformly dispersed Pb doped  $\text{Al}_2\text{O}_3$  nanoparticles is obtained. CH-Pb doped  $\text{Al}_2\text{O}_3$  hybrid nanobiocomposite films have been fabricated by uniformly dispersing 10  $\mu\text{L}$  solution of CH-Pb doped  $\text{Al}_2\text{O}_3$  for 12 h. These solution cast CH-Pb doped  $\text{Al}_2\text{O}_3$  hybrid nanobiocomposite films are washed repeatedly with deionized water to remove any unbound particles.

#### **Immobilization of Ur onto CH-Pb doped $\text{Al}_2\text{O}_3$ nanobiocomposite film**

10  $\mu\text{L}$  of bienzyme solution containing Ur (10 mg/ml) [prepared in phosphate buffer (0.1mol), pH 7] is immobilized onto CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode. The Ur/CH-Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite/gold bioelectrodes are kept undisturbed for about 12 h at  $4^\circ\text{C}$ . Finally, the dry bioelectrode is immersed in



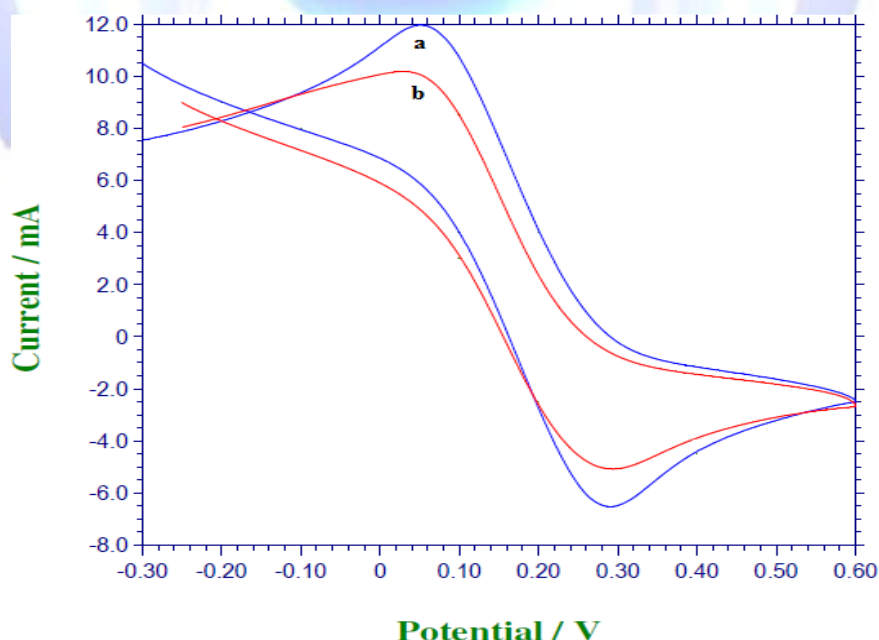
50 mM PBS(pH 7.0) in order to wash out any unbound enzymes from the electrode surface.

## Results and Discussions:

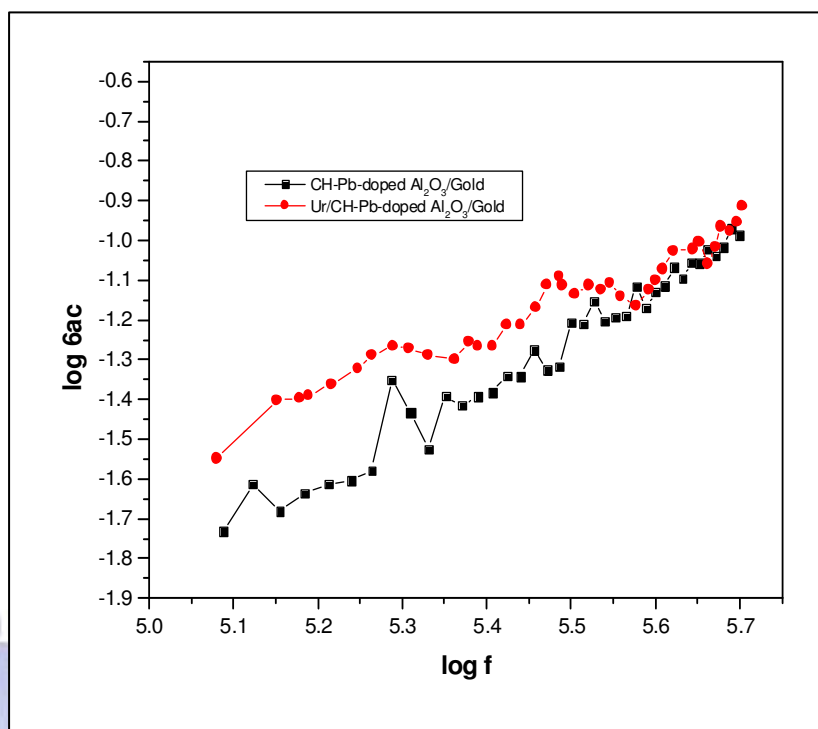
### Cyclic Voltammetry Studies

**Figure. 1.** Shows the cyclic voltammograms response of the prepared CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode and Ur/Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite electrode recorded in KCL solution containing 5M  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  in the potential range of -0.3 to 0.6 V at 10mV/s rate. The magnitude of the current response for CH-Pb doped  $\text{Al}_2\text{O}_3$  electrode increases in comparison to that of Ur/CH-Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite electrode. These results suggest that the presence of Pb doped  $\text{Al}_2\text{O}_3$  nanoparticles results in increased electroactive surface area of electrode resulting in enhanced electron transport between electrolyte medium and the electrode. However, magnitude of the current response decreases after immobilization of urease onto CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode. This may perhaps be due to insulating characteristics of urease.

Electrochemical impedance spectroscopy (EIS) can provide useful information on the impedance changes of the electrode surface during the fabrication process. **Figure. 2.** Shows the variation of ac conductivity of the prepared CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode and Ur/Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite electrode recorded in KCL solution containing 5M  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  in log-log scale. In the Figure, two different curves of CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode and Ur/Pb doped  $\text{Al}_2\text{O}_3$  nanobiocomposite electrodes frequency appears to merge at high frequency side. The ac conductivity increases with immobilization of urease on the surface of CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode.



**Figure. 1-** Cyclic voltammograms of (a) CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold electrode (b) Ur/CH-Pb doped  $\text{Al}_2\text{O}_3$ /gold bioelectrode in KCL (0.1m) containing  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  at  $10\text{mVs}^{-1}$



**Figure. 2-** Ac conductivity of Pb doped CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/goldelectrode; Ur/ CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/goldnanobiocomposite electrode in KCL (0.1m) containing [Fe(CN)<sub>6</sub>]<sup>3-/4-</sup>

## Conclusions:

In this summary CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/goldnanoparticles was synthesized by sol-gel method. Finally results of our research work lead to design new biosensor for determination of urea. The thin film of the prepared CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/gold/gold electrode and Ur/CH–CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/goldnanobiocomposite electrode is form by physical adsorption. Ur immobilized on CH–CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/goldnanobiocomposite electrode was studied by cyclic voltammtery which confirm the immobilization. And electrochemical impedance spectroscopy shows the ac conductivity increases with immobilization of urease on the surface of CH–Pb doped Al<sub>2</sub>O<sub>3</sub>/gold electrode.

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## References:

- [1] P. Lin, F. Yan, Adv Mater 24 (2012) 34-51.
- [2] HU. Khan, J. Jang, J.J. Kim, W. Knoll, J Am Chem Soc 133 (2011) 2170-2176.
- [3] T. Vo-Dinh, B.M. Cullum, D.L. Stokes, Sens Actuators B 74(2001) 2-11.
- [4] T. Haruyama, Adv Drug Deliv Rev 55 (2003) 393-401.





- [5] **KK. Jain KK**, Expert Rev MolDiagn 3 (2003) 153-161.
- [6] **G. Dhawan, G. Sumana, B.D. Malhotra**, Biochem. Engin.J. (2008).
- [7] **R. Singhal, A. Gambhir, M. K. Pandey, S. Annapoorni, B.D. Malhotra**, Biosens. Bioelectron. 17 (2002) 697-703.
- [8] **Rajesh, V. Bisht, W. Takashima, K. Kaneto**, Biomaterials 26 (2005) 3683-3690.
- [9] **J.V.D. Melo, S. Cosnier, C. Mousty, C. Martelet, N.J. Renault**, Anal. Chem. 74 (2002) 4037-4043.
- [10] **S. Tembe, M. Karve, S. Inamdar, S. Haram, J. Melo, SF. D'Souza**, Anal Biochem 349(2006)72-7.
- [11] **SD. Kumar, AV. Kulkarni, RG. Dhaneshwar, SF. D'Souza**, Bioelectrochem Bioenergetics 27 (1992)153-60.
- [12] **SD. Kumar, AV. Kulkarni, RG. Dhaneshwar, SF. D'Souza**, Bioelectrochem Bioenergetics 34 (1994)195-8.
- [13] **J. Kumar, SK. Jha, SF. D'Souza**, BiosensBioelectron 21 (2006)2100-5.
- [14] **KR. Rogers, CL. Gerlach**, Environ Sci Tech News (1999)500A-6A.
- [15] **K. Shanmugam, S. Subrahmanyam, SV. Tarakad, N. Kodandapani, D'Souza** Anal Sci 17 (2001)1369-74.
- [16] **SF. D'Souza**, Microbial biosensors. BiosensBioelectron 16 (200)337-53.
- [17] **SF. D'Souza**, ApplBiochemBiotechnol 96 (2001)225-38.
- [18] **LC. Shriver-Lake, B. Donner, R. Edelstein, K. Breslin, SK. Bhatia, FS.Ligler**, BiosensBioelectron 12 (1997)1101-6.
- [20] **JV. de Melo, AP. Soldatkin, C. Martelet, N. Jaffrezic-Renault, S. Cosnier**, BiosensBioelectron 18 (2003)345-51.

